

## Colony PCR-Phusion

- 33  $\mu$ l dH<sub>2</sub>O
- 10  $\mu$ l homemade 5X buffer Phusion buffer
- 5  $\mu$ l dNTPs (2.5 mM each)
- 1  $\mu$ l 5' primer
- 1  $\mu$ l 3' primer
- 1  $\mu$ l Phusion polymerase

This recipe is enough for two colony PCR reactions.

Works well with this PCR program

Colony program

- a) 94°C denaturation: 5 minutes
- b) 94°C denaturation: 30 seconds
- c) 55°C annealing: 30 seconds
- d) 72°C extension: 15 sec/kb

repeat b-d 30-36 times

- e) 72°C final extension: 3 minutes

Pick a decent sized colony with a P200 pipette tip. Touch to an agar plate with appropriate antibiotic, put tip into a PCR tube or well of a 96-well PCR plate containing 24.5  $\mu$ l of PCR mix and leave. Repeat with desired number of colonies. For positive control, add 0.5  $\mu$ l of an appropriate template, for negative control do not add anything.

Shake tips in PCR mix vigorously, remove tips and run on colony PCR program.