

Colony PCR recipe

- 37 μ l dH₂O
- 5 μ l 10X Pfu buffer (see Pfu purification protocol)
- 5 μ l dNTPs (2.5 mM each)
- 1 μ l 5' primer
- 1 μ l 3' primer
- 0.5 μ l Pfu DNA polymerase
- 0.5 μ l Taq DNA polymerase

Works well with this PCR program

- a) 94°C denaturation: 5 minutes
 - b) 94°C denaturation: 30 seconds
 - c) 55°C annealing: 30 seconds
 - d) 72°C extension: 1 min/kb
- repeat b-d 35 times
- e) 72°C final extension: 3 minutes

Pick a decent sized colony with a P200 pipette tip. Touch to an agar plate, put tip into a PCR tube or well of a 96-well PCR plate containing 24.5 μ l of PCR mix and leave. Repeat with desired number of colonies. For positive control, add 0.5 μ l of an appropriate template, for negative control do not add anything.

Shake tips in PCR mix vigorously, remove tips and run on colony PCR program.